

Modeling Alzheimer's disease and other proteopathies in vivo: Is seeding the key?

Review Article

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Summary. Protein misfolding and aberrant polymerization are salient features of virtually all central neurodegenerative disorders, including Alzheimer's disease (AD), Parkinson's disease, triplet repeat disorders, tauopathies, and prion diseases. In many instances, a single amino acid change can predispose to disease by increasing the production and/or changing the biophysical properties of a specific protein. Possible pathogenic similarities among the cerebral proteopathies suggest that therapeutic agents interfering with the proteopathic cascade might be effective against a wide range of diseases. However, testing compounds preclinically will require disease-relevant animal models. Numerous transgenic mouse models of β -amyloidosis, tauopathy, and other aspects of AD have now been produced, but none of the existing models fully recapitulates the pathology of AD. In an attempt to more faithfully replicate the human disease, we infused dilute AD-brain extracts into Tg2576 mice at 3-months of age (i.e. 5–6 months prior to the usual onset of β -amyloid deposition). We found that intracerebral infusion of AD brain extracts results in: 1) Premature deposition of β -amyloid in eight month-old, β -amyloid precursor protein (β APP)-transgenic mice (Kane et al., 2000); 2) augmented amyloid load in the injected hemisphere of 15 month-old transgenic mice; 3) evidence for the spread of pathology to other brain areas, possibly by neuronal transport mechanisms; and 4) tau hyperphosphorylation (but not neurofibrillary pathology) in axons passing through the injection site. The seeding of β -amyloid *in vivo* by AD brain extracts suggests pathogenic similarities between β -amyloidoses such as AD and other cerebral proteopathies such as the prionoses, and could provide a new model for studying the proteopathic cascade and its neuronal consequences in neurodegenerative diseases.

Keywords: Alzheimer's disease – Amyloid – Conformational disease – Huntington's disease – Parkinson's disease – Proteopathy – Senile plaque – Tau – Transgenic

Alzheimer's disease: Pathophysiology and relationship with other proteopathies

Research on Alzheimer's disease has burgeoned since the discovery in the 1970's and 1980's that basal forebrain neurons containing the neurotransmitter acetylcholine appear to be especially vulnerable to the disease process (Bowen et al., 1976; Davies and Maloney, 1976; Whitehouse et al., 1982; Geula and Mesulam, 1999). The "cholinergic hypothesis" spawned a wealth of basic research on all facets of AD, and led to the development of the first generation of approved therapies for AD in the United States (Giacobini and Becker, 1994). One outcome of this research was the reaffirmation that more than just the basal forebrain cholinergic system degenerates in AD, including neurons using various monoamines, peptides and amino acids as neurotransmitters (Hedera and Whitehouse, 1994; Sisodia et al., 1995; see also Martin (1999) for a critical review). The result of the degenerative process is widespread, but selective, neurochemical alterations and synaptic deafferentation, which contribute importantly to the dementia that characterizes AD (Terry et al., 1991; Hof and Morrison, 1999). Although therapeutic strategies for restoring transmitter function can be expected to improve cognitive decline to some degree when sufficient neuronal substrate remains, retarding the underlying disease process holds the best hope for long-term, effective treatment. Hence, many researchers in the field are returning to senile plaques and

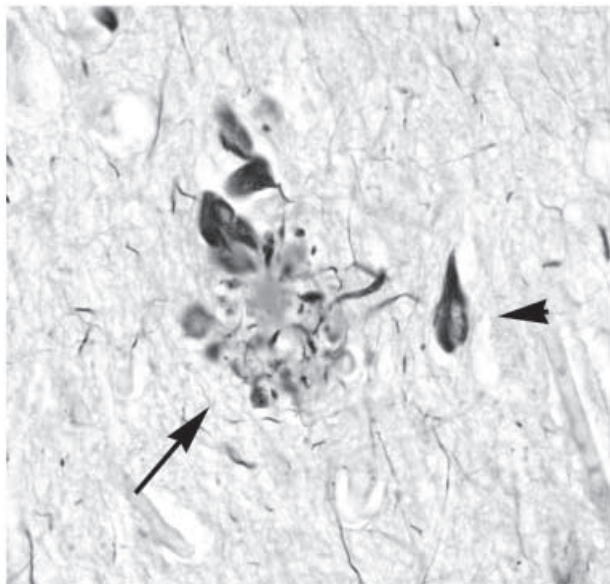


Fig. 1. Senile plaque (arrow) and neurofibrillary tangle (arrowhead) in the hippocampal formation of a patient who had died of AD. The cores of senile plaques are formed primarily of aggregated, fibrillar $A\beta$, whereas neurofibrillary tangles are bundles of paired helical filaments that are composed of abnormally polymerized tau protein. Note the numerous dark, enlarged, abnormal neurites that surround the light-gray plaque core. Naoumenko-Feigen silver stain, periodic acid-Schiff (PAS) counterstain

neurofibrillary tangles (the canonical neuropathological hallmarks of AD; Fig. 1) as the main source of insight into the origins of AD and into potential disease-modifying therapies.

The most salient feature of senile plaques and neurofibrillary tangles is the abnormal assembly and accumulation of particular proteins. The peptide $A\beta$ aggregates into senile plaques in the brain parenchyma, and also collects to a variable extent in the walls of cerebral blood vessels. Hyperphosphorylated forms of the microtubule-associated protein *tau* polymerize within neurons to form paired helical filaments (identified with the light microscope as neurofibrillary tangles). Exactly how $A\beta$ and tau contribute to disease pathogenesis is still disputed. With regard to $A\beta$, a growing school of thought holds that a non-fibrillar intermediate, such as an oligomeric form of the peptide, is responsible for cytopathology, and that plaques are just an obvious byproduct of this more insidious process (Klein et al., 2001; Walker & LeVine III, 2000b).

In the context of aging, specific mutations in the gene encoding β APP (the protein that spawns $A\beta$) result in autosomal dominant AD (Selkoe, 1999), as do mutations in the presenilins (St. George-Hyslop,

1999) (proteins that are involved in the carboxy-terminal gamma-site cleavage of $A\beta$ from β APP). Familial AD and the more common “sporadic” forms of AD are characterized by the same types of lesions, although such factors as age of onset and rate of progression, as well as various other known and unknown modifiers, can influence the degree and distribution of pathology. The involvement of $A\beta$ is constant, despite the phenotypic variability of AD. Indeed, all known risk factors for AD result in an increase in the production and/or deposition of $A\beta$ (Selkoe, 1999).

The participation of tau in the pathogenesis of AD also is indisputable, but which lesion initiates the Alzheimer’s disease process, the plaque or the tangle? The familial tauopathies (Mandelkow and Mandelkow, 1998; Spillantini and Goedert, 1998; Hardy et al., 1998; Lee and Trojanowski, 1999; Tolnay and Probst, 1999) suggest a tentative answer to this question. Certain mutations in tau result in dementing disorders characterized histologically by numerous neurofibrillary tangles, but β -amyloid deposits are usually absent (Buée et al., 2000). Thus, mutations in tau result in dementing disorders characterized pathologically only by the presence of tangles, but mutations in β APP engender *both* senile plaques and neurofibrillary tangles in Alzheimer’s disease. Interestingly, in AD, tau pathology tends to correlate better with dementia than does plaque load (Berg et al., 1993; Arriagada et al., 1998). Hence, it appears that tauopathy is sufficient to cause dementia, but the road to tau pathology in Alzheimer’s disease first passes through $A\beta$. In any event, both tau and $A\beta$ are legitimate therapeutic targets for altering the course of AD.

In fact, the misfolding and aberrant polymerization of specific proteins are common characteristics of an astonishing variety of diseases of the central nervous system and beyond (Walker and LeVine III, 2000a; Walker and LeVine III, 2000b). In many instances, a single amino acid change is sufficient to render a protein pathogenic (various posttranslational modifications as well as interactions with other proteins also can play a part). Understanding these proteopathic mechanisms is the next major frontier for biomedical research; to reach it, valid animal models will be needed. In the 1980’s and early 1990’s, the only available animal models of cerebral β -amyloidosis were aged dogs and nonhuman primates, which naturally produce β -amyloid (Walker et al., 1994; Cummings et al., 1996; Walker, 1997a,b; Walker and Cork, 1999; Walker, 2000). More recently, genetically modified

mice have been playing a growing part in establishing the significance of protein aggregation in neurodegenerative processes, in dissecting the pathogenic mechanisms involved, and in testing novel therapies.

Transgenic mouse models of Alzheimer pathology

After some early setbacks (Marx, 1992), the AD field has enjoyed a profusion of transgenic and/or knockout mice for a variety of AD-related proteins, such as β APP, tau, presenilins, and apolipoprotein E (for reviews see Walker, 1997a; Price et al., 1998; Sommer, 1998; Masliah, 1999). The outcome is a wealth of *in vivo* models of aspects of AD, but, to date, no mouse acquires the complete pathology of AD. β APP-transgenic mice, including some with plentiful, age-associated A β -deposition, have been particularly well-characterized (Walker, 1997a; Price et al., 1998; Sommer, 1998; Masliah, 1999; Callahan et al., 2001). The components of senile plaques in β APP-transgenic mice resemble in many ways those seen in humans (Games et al., 1995; Hsiao et al., 1996; Irizarry et al., 1997; Sturchler-Pierrat et al., 1997; Frautschy et al., 1998; Masliah, 1999; Stalder et al., 1999), except that neurofibrillary tangles have never been seen, either in the abnormal neurites that surround plaques or in neuronal somata.

Many groups have now developed mouse models of tauopathy that express different tau protein constructs (Götz et al., 1995; Brion et al., 1999; Ishihara et al., 1999; Spittaels et al., 1999; Duff et al., 2000; Lewis et al., 2000; Probst et al., 2000; Götz et al., 2001). Because hyperphosphorylation of tau is thought to contribute to its pathogenic state (Buée et al., 2000), mice expressing sundry transgenes that promote phosphorylation also have been generated (James et al., 1996; Brownlees et al., 1997; Ahljianian et al., 2000; Tesseur et al., 2000; Tesseur et al., 2000; Bian et al., 2002). Possibly the closest approximation to human tauopathy is a transgenic mouse model that expresses mutant (P301L) tau protein (Lewis et al., 2000; Götz et al., 2001), but there remain important differences between the murine and human lesions. In short, there is still no completely faithful transgenic mouse model of human-like tangles, but, as with β APP transgenics, the field is still young. As a possible means of stimulating more human-like pathology in mice, we have been investigating the intracerebral infusion of AD brain extract in β APP-transgenic mice. Our findings are briefly reviewed below.

The Tg2576 (Hsiao) mouse model of β -amyloidosis

Tg2576 mice overexpress the human β APP transgene (695 amino acid isoform) with the “Swedish” double mutation (K670N-M671L) driven by a hamster prion promoter (Hsiao et al., 1996). Protein expression with this construct is directed primarily to neurons and astrocytes, and in brain is approximately five times the expression of endogenous, mouse β APP. Although the expression of the transgene remains stable as the mice age, the presence of cerebral A β begins to increase dramatically around the age of nine months, in conjunction with the appearance of small, scattered senile plaques in certain brain regions (Hsiao et al., 1996; Callahan et al., 2001). By 24 months of age, amyloid burden can be pronounced (Fig. 2), although interindividual variability is high. Surprisingly, female Tg2576 mice deposit significantly more β -amyloid than do male mice (Callahan et al., 2001). Both plaque load measured histologically and the levels of the 40- and 42-amino acid versions of A β (A β 40 and A β 42) measured by ELISA are increased in the females. The effect of sex is most evident at around 15–19 months of age, when nearly three times as much cortical/hippocampal area is occupied by plaques in females than in males.

Seeding A β proteopathy in Tg2576 mice

If, as we suspect, the abnormal assembly of A β into multimeric species impels the neurodegenerative process, then it is imperative to understand how A β polymerization is initiated and sustained. Taking our cue from early studies of the prionoses (Prusiner et al., 1990; Prusiner et al., 1999), as well as primate studies of intracerebral AD homogenate injections (Baker et al., 1994), we hypothesized that A β aggregation might be induced, or “seeded”, in β APP-transgenic mice by material from diseased brains (Kane et al., 2000). To test this hypothesis, we infused dilute extracts of AD- and control brain homogenates into young (3-month-old), male Tg2576 mice. Autopsy-derived samples of human neocortex were homogenized, centrifuged to remove plaque cores, blood vessels and other debris, and the clear supernatant diluted to 1% final concentration (w/v). The resulting extracts from four AD cases, one young control, and one aged control case were then infused unilaterally into the dorsal hippocampus and overlying neocortex of Tg2576 and non-transgenic littermate control mice. In the month

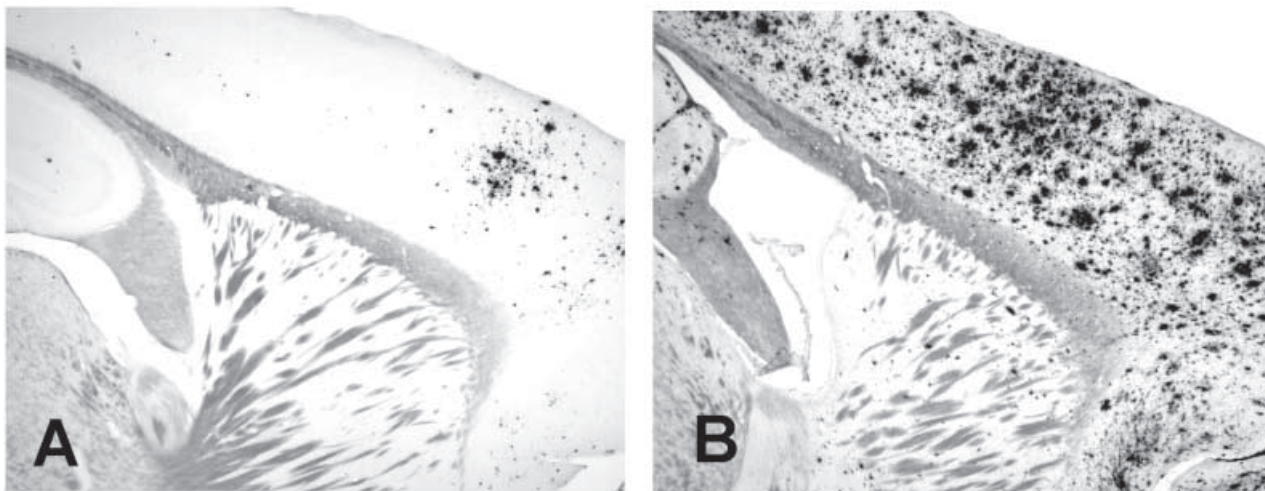


Fig. 2. β -Amyloid accumulation in 15-month-old (A) and in 24-month-old (B) Tg2576 mice. The amyloid deposits are mostly diffuse in nature, and are stained black. White matter pathways appear light gray in these black and white images. Note the massive accumulation of amyloid in the neocortex (top and right of the tissue section) at age 24 months. $A\beta$ -deposits also become increasingly numerous in subcortical structures of severely affected animals. Sagittal section through the anterior forebrain, rostral is to the right. Campbell-Gallyas silver stain

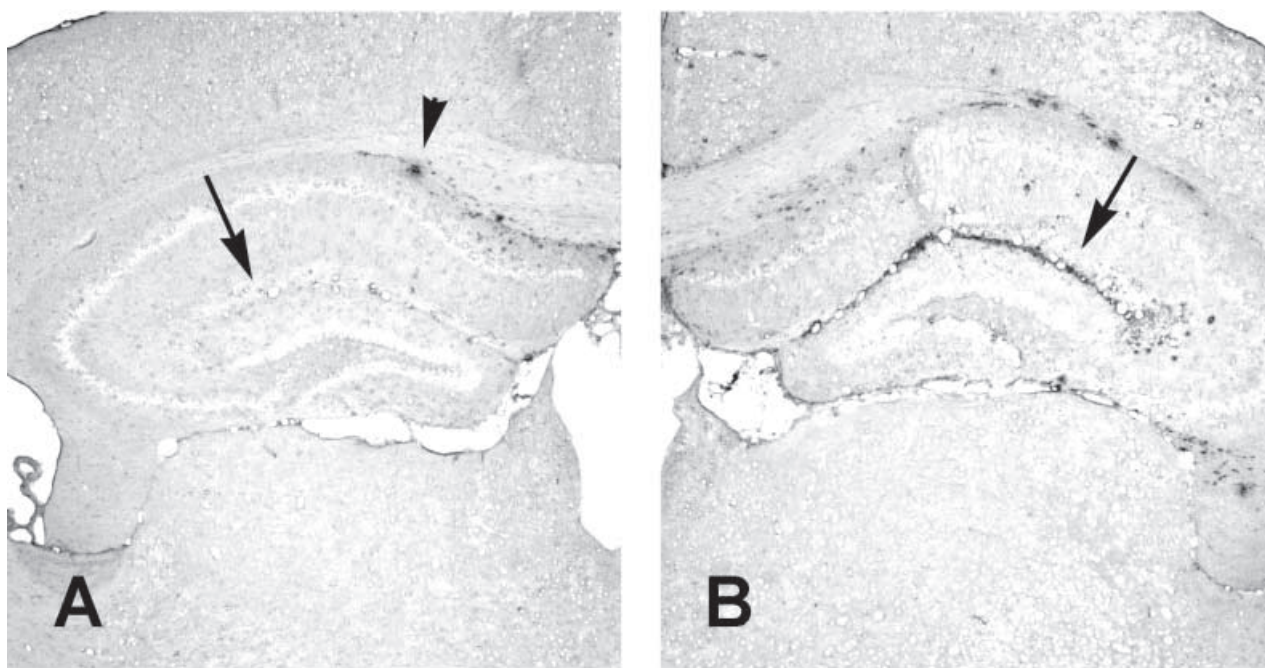


Fig. 3. Seeded, $A\beta$ -immunoreactive amyloid deposits in the AD extract-injected hemisphere (B) and in the non-injected hemisphere (A) of an 8-month-old Tg2576 mouse injected at three months of age. Note the heavy accumulation of $A\beta$ along the hippocampal fissure on the injected side (the fissure is indicated in each hemisphere by an arrow). Note also small patches of $A\beta$ -immunoreactivity extending along the ventral corpus callosum into the non-injected hemisphere (arrowhead denotes one deposit). The amyloid burden was approximately ten times greater in the injected hemisphere than in the non-injected hemisphere (see Kane et al., 2000). Coronal section, antibody 4G8 immunostain for $A\beta$

immediately following injection, there was no evidence of A β immunoreactivity. Five months later (i.e., at eight months of age) the transgenic mice that had been injected with AD brain homogenate had pronounced deposition of diffuse and compact A β deposits primarily in the injected hemisphere (Fig. 3). The non-transgenic control mice were negative, as were transgenic mice injected with the brain homogenate from a young, control human. Interestingly, homogenate from a non-demented aged human with mild β -amyloidosis stimulated a small amount of A β deposition in Tg2576 mice (Kane et al., 2000).

To assess the long-term effects of seeding, we recently studied mice one year post-infusion, at the age of 15 months. These mice had abundant β -amyloid deposition bilaterally (as expected in Tg2576 mice of this age) that was concentrated most heavily in the injected hemisphere. There was also evidence of hyperphosphorylated tau (antibody AT8)-immunoreactive axons in the surgery-damaged corpus callosum, particularly where this white matter pathway was beset by amyloid deposits (Walker et al., submitted). In the five-month seeded mice, we had observed, in the ipsilateral entorhinal cortex of some AD extract-injected mice, a small patch of A β -immunoreactive microglial cells that intermingled with neurons projecting to the extract-injection site (Walker et al., submitted). At one year post-injection, the ipsilateral entorhinal cortex contained a much greater plaque load than did the contralateral entorhinal cortex, suggesting that axonal transport mechanisms contribute to the spread of the seeded pathology.

Our data thus show that β -amyloid deposition can be actuated by exogenous material. Studies are currently in progress to identify the necessary factors in the brain extract for seeding A β *in vivo*. This seeding paradigm can provide insights into the early pathogenesis of β -amyloidosis and related pathologies, and may also reveal important commonalities in the mechanisms whereby diverse proteopathies are instigated and propagated in the brain.

References

- Ahlijanian MK, Barrezueta NX, Williams RD, Jakowski A, Kowsz KP, McCarthy S, Coskran T, Carlo A, Seymour PA, Burkhardt JE, Nelson RB, McNeish JD (2000) Hyperphosphorylated tau and neurofilament and cytoskeletal disruptions in mice over-expressing human p25, an activator of cdk5. *Proc Natl Acad Sci USA* 97: 2910–2915
- Arriagada PV, Growdon JH, Hedley-Whyte T, Human BT (1998) Neurofibrillary tangles but not senile plaques parallel duration and severity of Alzheimer's disease. *Neurology* 42: 631–639
- Baker HF, Ridley RM, Duchon LW, Crow TJ, Bruton CJ (1994) Induction of beta (A4)-amyloid in primates by injection of Alzheimer's disease brain homogenate. Comparison with transmission of spongiform encephalopathy. *Molec Neurobiol* 8: 25–39
- Berg L, McKeel DW, Miller JF, Baty J, Morris JC (1993) Neuropathological indexes of Alzheimer's disease in demented and nondemented persons aged 80 years and older. *Arch Neurol* 50: 349–358
- Bian F, Nath R, Sobocinski G, Booher RN, Lipinski WJ, Callahan MJ, Pack A, Wang KK-W, Walker LC (2002) Axonopathy, tau abnormalities, and dyskinesia, but no neurofibrillary tangles in p25-transgenic mice. *J Comp Neurol* 446: 257–266
- Bowen DM, Smith CB, White P, Davison AN (1976) Neurotransmitter-related enzymes and indices of hypoxia in senile dementia and other abiotrophies. *Brain* 99: 459–496
- Brion JP, Tremp G, Octave JN (1999) Transgenic expression of the shortest human tau affects its compartmentalization and its phosphorylation as in the pretangle stage of Alzheimer's disease. *Am J Pathol* 154: 255–270
- Brownlee J, Irving NG, Brion JP, Gibb BJ, Wagner U, Woodgett J, Miller CC (1997) Tau phosphorylation in transgenic mice expressing glycogen synthase kinase-3beta transgenes. *Neuroreport* 8: 3251–3255
- Buée L, Bussière T, Buée-Scherrer V, Delacourte A, Hof PR (2000) Tau protein isoforms, phosphorylation and role in neurodegenerative disorders. *Brain Res Rev* 33: 95–130
- Callahan MJ, Lipinski WJ, Bian F, Durham RA, Pack A, Walker LC (2001) Augmented senile plaque load in aged female β -amyloid precursor protein-transgenic mice. *Am J Pathol* 158: 1173–1177
- Cummings BJ, Head E, Ruehl W, Milgram NW, Cotman CW (1996) The canine as an animal model of human aging and dementia. *Neurobiol Aging* 17: 259–268
- Davies P, Maloney AJ (1976) Selective loss of cholinergic neurons in Alzheimer's disease. *Lancet* ii: 1403
- Duff K, Knight H, Refolo LM, Sanders S, Yu X, Picciano M, Malester B, Hutton M, Adamson J, Goeder M, Davies P (2000) Characterization of pathology in transgenic mice over-expressing human genomic and cDNA tau transgenes. *Neurobiol Dis* 7: 87–98
- Frautschy SA, Yang F, Irrizarry M, Hyman B, Saido TC, Hsiao K, Cole GM (1998) Microglial response to amyloid plaques in APPsw transgenic mice. *Am J Pathol* 152: 307–317
- Games D, Adams D, Alessandrini R, Barbour R, Berthelette P, Blackwell C, Carr T, Clemens J, Donaldson T, Gillespie F, et al (1995) Alzheimer-type neuropathology in transgenic mice. *Nature* 373: 523–527
- Geula C, Mesulam M-M (1999) Cholinergic systems in *Alzheimer disease*. In: Terry RD, Katzman R, Bick KL, Sisodia SS (eds) *Alzheimer disease*. Lippincott Williams & Wilkins, Philadelphia, pp 269–292
- Giacobini E, Becker R (1994) Development of drugs for Alzheimer therapy: a decade of progress. In: Giacobini E, Becker R (eds) *Alzheimer disease: therapeutic strategies*. Birkhäuser, Boston, pp 1–7
- Götz J, Chen F, Barmettler R, Nitsch RM (2001) Tau filament formation in transgenic mice expressing P301L tau. *J Biol Chem* 276: 529–534
- Götz J, Probst A, Spillantini MG, Schäfer T, Jakes R, Bürki K, Goedert M (1995) Somatodendritic localization and hyperphosphorylation of tau protein in transgenic mice expressing the longest human brain tau isoform. *EMBO J* 14: 1304–1313

- Hardy J, Duff K, Hardy KG, Perez-Tur J, Hutton M (1998) Genetic dissection of Alzheimer's disease and related dementias: amyloid and its relationship to tau. *Nature Neurosci* 1: 355–358
- Hedera P, Whitehouse PJ (1994) Neurotransmitters in neurodegeneration. In: Calne DB (ed) *Neurodegenerative disease*. W.B. Saunders Company, Philadelphia, pp 97–118
- Hof PR, Morrison JH (1999) The cellular basis of cortical disconnection in Alzheimer disease and related dementing conditions. In: Terry RD, Katzman R, Bick KL, Sisodia SS (eds) *Alzheimer disease*. Lippincott Williams & Wilkins, Philadelphia, pp 207–232
- Hsiao K, Chapman P, Nilsen S, Eckman C, Harigaya Y, Younkin S, Yang F, Cole G (1996) Correlative memory deficits, A beta elevation, and amyloid plaques in transgenic mice. *Science* 274: 99–102
- Irizarry MC, Soriano F, McNamara M, Page KJ, Schenk D, Games D, Hyman BT (1997) Abeta deposition is associated with neuropil changes, but not with overt neuronal loss in the human amyloid precursor protein V717F (PDAPP) transgenic mouse. *J Neurosci* 17: 7053–7059
- Ishihara T, Hong M, Zhang B, Nakagawa Y, Lee MK, Trojanowski JQ, Lee VM (1999) Age-dependent emergence and progression of a tauopathy in transgenic mice overexpressing the shortest human isoform. *Neuron* 24: 751–762
- James ND, Davis DR, Sinden J, Hanger DP, Brion JP, Miller CC, Rosenberg MP, Anderton BH, Probst F (1996) Neurodegenerative changes including altered tau phosphorylation and neurofilament immunoreactivity in mice transgenic for the serine/threonine kinase Mos. *Neurobiol Aging* 17: 235–241
- Kane MD, Lipinski WJ, Callahan MJ, Bian F, Durham RA, Schwarz RD, Roher AE, Walker LC (2000) Evidence for seeding of beta-amyloid by intracerebral infusion of Alzheimer brain extracts in beta-amyloid precursor protein-transgenic mice. *J Neurosci* 20: 3606–3611
- Klein WL, Krafft GA, Finch CE (2001) Targeting small A β oligomers: the solution to an Alzheimer's disease conundrum? *TRENDS Neurosci* 24: 219–234
- Lee VM, Trojanowski JQ (1999) Neurodegenerative tauopathies: human disease and transgenic mouse models. *Neuron* 24: 507–510
- Lewis J, McGowan E, Rockwood J, Melrose H, Nacharaju P, Van Slegtenhorst M, Gwinn-Hardy K, Paul Murphy M, Baker M, Yu X, Duff K, Hardy J, Corral A, Lin WL, Yen SH, Dickson DW, Davies P, Hutton M (2000) Neurofibrillary tangles, amyotrophy and progressive motor disturbance in mice expressing mutant (P301L) tau protein. *Nature Gen* 25: 402–405
- Mandelkow E-M, Mandelkow E (1998) Tau in Alzheimer's disease. *Trends Cell Biol* 8: 425–427
- Martin JB (1999) Alzheimer disease: neurochemical aspects. In: Terry RD, Katzman R, Bick KL, Sisodia SS (eds) *Alzheimer disease*. Lippincott Williams & Wilkins, Philadelphia, pp 263–267
- Marx J (1992) Major setback for Alzheimer's models. *Science* 255: 1200–1202
- Masliah E (1999) Transgenic animal models of Alzheimer disease. In: Terry RD, Katzman R, Bick KL, Sisodia SS (eds) *Alzheimer disease*. Lippincott Williams & Wilkins, Philadelphia, pp 245–261
- Price DL, Sisodia SS, Borchelt DR (1998) Genetic neurodegenerative diseases: The human illness and transgenic models. *Science* 282: 1079–1083
- Probst A, Götz J, Wiederhold KH, Tolnay M, Mistl C, Jaton AL, Hong M, Ishihara T, Lee VM, Trojanowski JQ, Jakes R, Crother RA, Spillantini MG, Bürki K, Goedert M (2000) Axonopathy and amyotrophy in mice transgenic for human four-repeat tau protein. *Acta Neuropathol* 99: 469–481
- Prusiner SB, Safar J, Cohen FE, De Armond SJ (1999) The prion diseases. In: Terry RD, Katzman R, Bick KL, Sisodia SS (eds) *Alzheimer disease*. Lippincott Williams & Wilkins, Philadelphia, pp 161–179
- Prusiner SB, Scott M, Foster D, Pan KM, Groth D, Mirenda C, Torchia M, Yang SL, Serban D, Carlson GA (1990) Transgenic studies implicate interactions between homologous PrP isoforms in scrapie prion replication. *Cell* 63: 673–686
- Selkoe DJ (1999) Biology of beta-amyloid precursor protein and the mechanism of Alzheimer disease. In: Terry RD, Katzman R, Bick KL, Sisodia SS (eds) *Alzheimer disease*. Lippincott Williams & Wilkins, Philadelphia, pp 293–310
- Sisodia SS, Martin LJ, Walker LC, Borchelt DR, Price DL (1995) Cellular and molecular biology of Alzheimer's disease and animal models. *Neurodegen Dis* 5: 59–68
- Sommer B (1998) Recent advances in transgenic model development for Alzheimer's disease. *Exp Opin Invest Drugs* 7: 2017–2025
- Spillantini MG, Goedert M (1998) Tau protein pathology in neurodegenerative diseases. *TRENDS Neurosci* 21: 428–433
- Spittaels K, Van den Haute C, Can Dorpe J, Bruynseels K, Vandezande K, Laenen I, Geerts H, Mercken M, Van Lommel A, Loos R, Van Leuven F (1999) Prominent axonopathy in the brain and spinal cord of transgenic mice overexpressing four-repeat human tau protein. *Am J Pathol* 155: 2153–2165
- St. George-Hyslop PH (1999) Molecular Genetics of Alzheimer disease. In: Terry RD, Katzman R, Bick KL, Sisodia SS (eds) *Alzheimer disease*. Lippincott Williams & Wilkins, Philadelphia, pp 311–326
- Stalder M, Phinney A, Probst A, Sommer B, Staufenbiel M, Jucker M (1999) Association of microglia with amyloid plaques in brains of APP23 transgenic mice. *Am J Pathol* 154: 1673–1684
- Sturchler-Pierrat C, Abramowski D, Duke M, Wiederhold KH, Mistl C, Rothacher S, Ledermann B, Bürki K, Paganetti PA, Waridel C, Calhoun ME, Jucker M, Probst A, Staufenbiel M, Sommer B (1997) Two amyloid precursor protein transgenic mouse models with Alzheimer disease-like pathology. *Proc Natl Acad Sci USA* 94: 13287–13292
- Terry RD, Masliah E, Salmon DP, Butters N, DeTeresa R, Hill R, Hansen LA, Katzman R (1991) Physical basis of cognitive alterations in Alzheimer's disease: Synapse loss is the major correlate of cognitive impairment. *Ann Neurol* 30: 572–580
- Tesseur I, Van Dorpe J, Bruynseels K, Bronfman F, Sciôt R, Van Lommel A, Van Leuven F (2000) Prominent axonopathy and disruption of axonal transport in transgenic mice expressing human apolipoprotein neurons of brain and spinal cord. *Am J Pathol* 157: 1494–1510
- Tesseur I, Van Dorpe J, Spittaels K, Van den Haute C, Moerchars D, Van Leuven F (2000) Expression of human apolipoprotein E4 in neurons causes hyperphosphorylation of protein tau in the brains of transgenic mice. *Am J Pathol* 156: 951–964
- Tolnay M, Probst A (1999) Tau protein pathology in Alzheimer's disease and related disorders. *Neuropath Appl Neurobiol* 25: 171–187
- Walker LC (1997a) Animal models and Alzheimer's disease. *Alzheimer's Dis Invest Drugs (ID) Res Alert* 2: 133–139
- Walker LC (1997b) Animal models of cerebral β -amyloid angiopathy. *Brain Res Rev* 25: 70–84
- Walker LC (2000) Cerebral amyloid angiopathy in aged dogs and nonhuman primates. In: Verbeek MM, Vinters HV (eds) *Cerebral amyloid angiopathy in Alzheimer's disease and related disorders*. Kluwer Academic Publishers, Boston, pp 313–324
- Walker LC, Callahan MC, Bian F, Durham RA, Roher AE, Lipinski WJ (submitted) Exogenous induction of cerebral β -amyloidosis in β APP-transgenic mice.
- Walker LC, Cork LC (1999) The neurobiology of aging in nonhuman primates. In: Terry RD, Katzman R, Bick KL, Sisodia

- SS (eds) Alzheimer disease. Lippincott Williams & Wilkins, Philadelphia, pp 233–243
- Walker LC, LeVine III H (2000a) The cerebral proteopathies. *Neurobiol Aging* 21: 559–561
- Walker LC, LeVine III H (2000b) The cerebral proteopathies: Neurodegenerative disorders of protein conformation and assembly. *Molec Neurobiol* 21: 83–95
- Walker LC, Price DL, Voytko ML, Schenk DB (1994) Labeling of cerebral amyloid in vivo with a monoclonal antibody. *J Neuropath Exp Neurol* 53: 377–383
- Whitehouse PJ, Price DL, Struble RG, Clark AW, Coyle JT, DeLong MR (1982) Alzheimer's disease and senile dementia: loss of neurons in the basal forebrain. *Science* 215: 1237–1239
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